	Domain	Sample	Analytical techniques	Time limit			Special Features
				Optimal timeframe	Acceptable delay (Results given with reservations)	Deadline exceeded (Sample rejected)	
Internal parasites	▶ **Coproscopic examination: qualitative method, quantitative method, or combined method[1]**	▪ Faeces. ▪ Rectal contents ▪ Intestinal contents	* Zinc sulfate flotation * NaCl flotation	≤ 72 h	≤ 7 days	> 7 days	Quantity > 10 g
	▶ **Cryptosporidium**		* ELISA or sucrose flotation	≤ 72 h	3 to 10 days	> 10 days	Quantity > 10 g
	▶ ***Giardia* spp.**		* Immunochromatographic test * PCR	≤ 72 h	3 to 10 days	> 10 days	Quantity > 10 g
	▶ **Lungworm larvae**	▪ Faeces ▪ Intestinal contents ▪ ATT liquid ▪ Lung	* Baermann's method	≤ 24 h	2 to 3 days	> 3 days	Quantity of feces > 10 g
	▶ **Qualitative parasitological examination**	▪ Organs	* Direct examination	≤ 48 h	2 to 10 days	> 10 days	
External parasite	▶ **Mites (sarcoptic, demodectic or auricular scabies)**	▪ Deep skin scraping product for the affected areas ▪ Ear swabbing ▪ Pulled hairs (with bulbs)	* Direct examination	≤ 72 h	/	/	

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Mycology	► **Dermatophytes (ringworm)**	▪ Hairs plucked from the edge of the active lesion ▪ Shave the area and then gently scrape the edges of the lesion.	* Direct examination and culture	≤ 72 h	/	/	

[1]Selection of the appropriate coproscopic analysis method based on the main contexts

Combined method (quantitative and qualitative)

- **Objective :** ensure parasite surveillance through exhaustive coproscopic examination and broad-spectrum, seasonal monitoring, in order to avoid any unnecessary treatment.

Quantitative method

- **Objective :** clinical diagnosis (one-time pathological event).
  - **Coccidia**
  - **Digestive strongyles**
  - ***Strongyloides***

Qualitative method

- **Objective :** clinical diagnosis (one-time pathological event).
- **Trematodes**
- **Cestodes**

Recommendations :

- To ensure the reliability of the results, provide a representative sample that has not been altered during transport and storage.
- The time between sampling and analysis must not exceed 7 days.
- Fecal pooling is only useful for certain investigations (shedding intensity, treatment efficacy). It should be avoided for parasites with low excretion rates (*Fasciola, Dicrocoelium*, etc.). Results cannot be extrapolated to individual samples. For greater reliability, pooling can be prepared in the laboratory using a maximum of 5 samples.
- Collect fecal matter directly from the rectum or intestine (in deceased animals). Avoid collecting feces from the environment to limit contamination. If this is not possible (e.g., young animals), collect only fresh feces from clean bedding, without scraping the ground.
- For external parasites and dermatophytes: sterile tubes, aluminum foil, or Petri dishes for transport to the laboratory. Rapid transport to the laboratory.

Sample storage :

- The sample must be placed in a sealed vial.
- Refrigerate quickly according to shipping conditions and outside temperature.
- Send to the laboratory as soon as possible. Do not freeze the sample.

Transport packaging:

- Triple waterproof packaging is recommended.

Rejection criteria

- Insufficient quantity.
- Acceptance deadline exceeded.
- Frozen sample.